

Snakebite-Induced Acute Kidney Injury in Latin America

Fábia M. Oliveira Pinho, MD, PhD,* Luis Yu, MD, PhD,†
and Emmanuel A. Burdmann, MD, PhD‡

Summary: There are 4 genera of venomous snakes in Latin America: *Bothrops*, *Crotalus*, *Lachesis*, and *Micrurus*. Acute kidney injury (AKI) has been reported consistently after *Bothrops* and *Crotalus* envenomations. In fact, these 2 genera of snakes are responsible, along with the Russell's viper, for the majority of cases of snakebite-induced AKI reported worldwide. Although the *Bothrops* snakes are the leading cause of venomous snakebites in Latin America, the absolute number of AKI cases seen after *Bothrops* and *Crotalus* snakebites is similar. In this article the main characteristics of *Bothrops* and *Crotalus* snakes and their venoms, the clinical picture, and the pattern of accidents, risk factors, and mechanisms of renal injury are reviewed.

Semin Nephrol 28:354-362 © 2008 Elsevier Inc. All rights reserved.

Keywords: Snakebite, acute kidney injury, Latin America, *Bothrops*, *Crotalus*

There are around 3,000 species of snakes, of which approximately 19% are venomous. The World Health Organization estimates the occurrence of 2,682,500 accidents by poisonous snakes per year worldwide, with 125,345 deaths and about 100,000 severe sequelae. Most of these accidents occur in tropical regions, where they represent a serious public health burden because of its incidence, morbidity, and mortality. Latin America is the third most affected area after Africa and Asia.^{1,2}

Snakebites are more common in the rainy seasons and are related to the increase of human activity in rural areas. The most affected group is 25- to 49-year-old men. Lower limbs are the most frequently injured site.

Venomous snakebite mortality rates vary in different regions of the world. In Asia, especially in India, Myanmar, and Malaysia, there are more than 2 million cases of snake envenomation per year, mostly by the *Vipera russelli* snake, with approximately 100,000 deaths.¹ In Nigeria, there are 600 cases per 100,000 inhabitants, with a mortality rate of 12%, and the *Echis* sp snakes are responsible for most of the accidents.³ In Australia, the incidence is 3 to 18 cases per 100,000 inhabitants,⁴ and *Pseudonaja* and *Notechis* account for most of the deaths. In Europe, the United States, and Canada, snakebite envenomation is relatively rare, with 15 to 30 fatal cases among 8,000 accidents per year.^{1,5}

Epidemiologic data on snakebite envenomation in Latin America are scarce. In Brazil, there are 20,000 accidents by venomous snakes per year, an incidence of 13.5 accidents/100,000 inhabitants, and a mortality rate of about 0.45%.⁶ *Bothrops* snakes (jararaca and jararacuçu) are responsible for 90.5%, *Crotalus* (South American rattlesnake) are responsible for 7.7%, *Lachesis* (surucucu and surucutinga) are responsible for 1.4%, and *Micrurus* (coral) are responsible for 0.4% of the cases when the type of serpent was identified. *Crotalus* (1.9%)

*Department of Medicine, Catholic University of Goiás, Goiânia, Brazil.

†ARF Group, Division of Nephrology, University of São Paulo, São Paulo, Brazil.

‡Division of Nephrology, São José do Rio Preto Medical School, São José do Rio Preto, São Paulo, Brazil.

Dr. Emmanuel A. Burdmann is partially supported by grants from the Foundation for the Support of Research in the State of São Paulo (Fundação de Amparo à Pesquisa do Estado de São Paulo, FAPESP) and from the National Council for Scientific and Technological Development (Conselho Nacional de Desenvolvimento Científico e Tecnológico, CNPq).

Address reprint requests to Fábila M. Oliveira Pinho, MD, R. 86, no. 115, Sala: 104, Setor Sul Goiânia-Goiás, Brazil 74.083-330. E-mail: pinhofabia@bol.com.br

0270-9295/08/\$ - see front matter

© 2008 Elsevier Inc. All rights reserved. doi:10.1016/j.semnephrol.2008.04.004

and *Bothrops* (0.3%) have the highest lethality rates.⁶

Acute kidney injury (AKI) is one of the main complications after snakebite envenomation and it is an important cause of mortality for these patients. Snakebite-induced renal injury has been reported with almost all venomous snakes, however, AKI is more frequent with the *Vipera russelli* in Asia and the *Bothrops* and *Crotalus* in South America.^{7,8}

AKI AFTER CROTALUS ENVENOMATION

Crotalus Snake

The South American rattlesnake belongs to the *Viperidae* family, *Crotalinae* subfamily, *Crotalus* genus, and is represented in Brazil by a single species, *Crotalus durissus*, distributed into 5 subspecies,⁹ of which *Crotalus durissus terrificus* and *Crotalus durissus collilineatus* are the most important ones.⁶

Crotalus durissus snakes are robust and may reach 1 meter in length.^{10,11} They are not very agile and are less aggressive than *Bothrops* snakes. Its most notable characteristic is the presence of a rattle at the end of its tail, which facilitates its identification.^{6,12} These snakes usually are found in open fields, dry areas, sand, rocks, and, rarely, along the ocean shore. They have vespertine and crepuscular habits, eat small rodents, and their most common predators are birds.^{6,12,13}

Crotalus venom is a complex combination of enzymes, toxins, and peptides.¹⁴ The main toxic components are crotoxin, crotamine, gyroxin, convulxin, and a thrombin-like enzyme.^{15,16} Crotoxin represents more than 50% of the proteins in the venom and is responsible for its high toxicity.¹⁶ It has neurotoxic,¹⁷ myotoxic,¹⁸⁻²¹ and

nephrotoxic activity.²²⁻²⁴ Consequently, *Crotalus* venom effects are multifactorial and the most important clinical manifestations are neurotoxicity, myotoxicity, nephrotoxicity, and coagulating activity.^{15,25-27}

Clinical Manifestations

The clinical picture of the *Crotalus* envenomation includes mild local and systemic manifestations that usually are severe. Eyelid ptosis, blurred and/or double vision, ophthalmoplegia, and facial muscle paralysis are manifestations of the venom neurotoxicity. In addition, myotoxicity provokes generalized rhabdomyolysis, which is manifested clinically by generalized myalgia and myoglobinuria. The coagulating action, caused by the thrombin-like enzyme, produces blood incoagulability and afibrinogenemia in 40% to 50% of patients, but bleeding is a rare manifestation.^{6,28}

AKI Prevalence

AKI is the major complication in patients surviving the initial venom effects and it is considered the main cause of death in these accidents.^{26,28-30} Although the *Crotalus* snakebite occurs 10 times less frequently than the *Bothrops* snakebite, the absolute number of AKI cases reported with both snake genera is similar,³¹ suggesting increased nephrotoxicity for *Crotalus* venom. The prevalence of AKI associated with *Crotalus* envenomation ranges from 10% to 29% (Table 1).^{28,32-35} To date, only the study by Pinho et al³⁵ was prospective, evaluating 100 consecutive patients and assessing sequentially their creatinine clearances. This study found the highest prevalence of AKI.³⁵

Table 1. AKI Prevalence After *Crotalus* Envenomation

Study	Type	n	AKI Diagnosis	% AKI
Pinto et al, 1987 ³²	Retrospective	114	Creatinine increase	18
Silveira and Nishioka, 1992 ³³	Retrospective	87	Urea and creatinine increase	18
Jorge and Ribeiro, 1992 ²⁸	Retrospective	249	Presence of oligoanuria	13
Bucarety et al, 2002 ³⁴	Prospective	31	Creatinine increase (children <15 y)	10
Pinho et al, 2005 ³⁵	Prospective	100	Creatinine clearance (<60 mL/min/1.73m ²)	29

Pathophysiology of AKI

Experimental and clinical studies suggest that the pathogenesis of *Crotalus* venom-induced AKI likely is related to rhabdomyolysis, renal vasoconstriction, and direct tubular cell toxicity.

In 1985, Azevedo-Marques et al¹⁸ showed clinically that the *Crotalus* venom causes rhabdomyolysis and myoglobinuria associated with AKI. Later, it was shown that the venom, more specifically crotoxin, induces systemic and selective muscle injury in skeletal muscle groups composed of type I and IIa oxidative fibers, which are rather vascularized and rich in myoglobin.^{19,21} An experimental study confirmed that sublethal doses of *Crotalus* venom cause early rhabdomyolysis associated with significant renal blood flow and glomerular filtration rate decrease, without systemic blood pressure reduction.³⁶

Kidneys are particularly vulnerable to toxins because of the high blood flow and the ability to concentrate substances in the urine.³¹ *Crotalus* venom is excreted predominantly through the kidneys and the toxic components have direct and indirect action on renal cells.^{27,36} Crotoxin is probably the most important component responsible for renal injury.^{22,23}

Other factors potentially associated with *Crotalus* venom-induced AKI such as shock, hypotension, hemolysis, sepsis, or use of nephrotoxic drugs have not been confirmed in clinical and experimental studies.³⁵⁻³⁷

Risk Factors for the Development of AKI After *Crotalus* Snakebite

Time Period for Antivenom Administration

One milliliter of *Crotalus* antivenom (CAV), which consists largely of immunoglobulin F(ab')₂ fragments, neutralizes 1.5 mg of *C durissus* venom.⁶ Circulating venom already is not observed at 1 hour after CAV administration, whereas CAV titer remains high for up to 24 hours after therapy.^{38,39} The amount of CAV to be administered currently is determined by the severity of the accident. The period between the snakebite and administration of the specific

antivenom is an important factor for the development of complications because the venom will remain active until it becomes neutralized.

We recently showed that a time interval greater than 2 hours between a *Crotalus* snakebite and administration of the specific antivenom increases the risk of AKI development 10-fold.³⁵ In a consistent way, experimental data disclosed that CAV administration was only effective in preventing renal proximal tubular injury when it was performed simultaneously with the addition of the venom.⁴⁰ Previous studies also have suggested a correlation between renal injury and the time interval between the snakebite and CAV administration, and it was observed that the longer it took for CAV administration the higher the risk for AKI.^{20,33,34}

These results disclose how important it is to decentralize the venomous snakebite treatment to allow early administration of CAV effective doses. Antivenom always should be available at health centers and emergency services of small communities rather than concentrated in reference centers or hospitals.

Rhabdomyolysis

Crotalus venom causes rhabdomyolysis with a significant increase of creatine phosphokinase (CK) serum levels. After a *Crotalus* snakebite, patient enzyme levels at admission greater than 2,000 UI/L were associated with a 12-fold increase of the risk of developing AKI.³⁵ The most effective prophylactic measure for the prevention of rhabdomyolysis-induced AKI is extracellular volume expansion with saline solution, associated with sodium bicarbonate and mannitol.^{41,42} This solution should be started as early as possible and maintained until myoglobinuria disappears. However, a recent prospective study failed to show the efficacy of this treatment in the prevention of AKI development after the *Crotalus* snakebite³⁵ despite a urinary pH of greater than 6.5, which is considered ideal for the prevention of myoglobin-induced renal injury.^{6,41,43,44} It is possible that this lack of protection was related to the delay to start this preventive maneuver or to the intensity of the muscle injury (median CK serum levels, 50,000 IU/L). Finally, it is possible that without

the protective maneuver the prevalence of AKI would be higher than observed.

Patient Age

In a retrospective study, Silveira and Nishioka³³ showed that older age was associated with a greater risk of developing AKI. They concluded that for patients older than 40 years of age, the presence of myalgia and neurotoxic facies were predictive factors of renal injury.

In contrast, our prospective study showed that children (<12 y) had a prevalence of *Crotalus* snakebite-induced AKI almost 3 times greater than adults.³⁵ The Health Ministry's recommendation for antivenom administration does not take into account the victims' age and therefore children and adults will receive similar amounts of antivenom, related only to the severity of the accident rather than their age.⁶ However, children have lower blood volume and less body surface, leading to a greater concentration of the venom and to more severe systemic actions.⁴⁵ These results strongly suggest that the recommendation for antivenom administration in children must be deeply reviewed.

Urinary Volume

The maintenance of a urinary flow of 30 to 40 mL/h is recommended for adults and 1 to 3 mL/kg/h for children to prevent AKI after *Crotalus* envenomation.^{6,43} In the study by Pinho et al,³⁵ diuresis greater than 90 mL/h at admission of *Crotalus* snakebite victims was protective against AKI development. A higher urinary flow may allow decreased exposure of renal tubular cells to myoglobin and to the venom, with consequent injury attenuation and prevention of tubular lumen obstruction by myoglobin cylinders and cellular debris.

Characteristics of AKI

AKI develops early after *Crotalus* envenomation, occurring within the first 24 to 48 hours after the accident. This suggests that direct venom nephrotoxicity may occur in the clinical setting as experimentally shown.^{6,26,35,46}

Dialysis was necessary in 24% of the cases in the study by Pinho et al.³⁵ Previous studies reported a greater need of dialysis, ranging

from 68% to 77% of patients. This discrepancy probably is related to the low sensitivity of AKI diagnostic criteria used in these studies, which had identified only cases of severe renal injury.^{26,33,47}

The high fractional sodium excretion values found in patients developing AKI after *Crotalus* envenomation suggest renal proximal tubule cell injury. In fact, histologic injury usually found in *Crotalus* snakebite victims is acute tubular necrosis, although cases of interstitial nephritis also have been reported.^{18,35,47-49}

Mortality rates reported for AKI after *Crotalus* envenomation range from 8% to 17%. In addition, the majority of the snakebite victims are young and previously healthy individuals.^{26,33,35,47}

AKI AFTER *BOTHROPS* ENVENOMATION

Bothrops Snake

Snakes of the *Bothrops* genus belong to the *Viperidae* family and *Crotalinae* subfamily. There are more than 30 species distributed from southern Mexico to Argentina and Brazil. The most important species are *Bothrops asper* in Central America and *Bothrops atrox*, *Bothrops erythromelas*, *Bothrops neuwiedi*, *Bothrops moojeni*, *Bothrops jararaca*, *Bothrops jararacussu*, and *Bothrops alternatus*, found in Brazil, especially in grassland regions (called *cerrados*) and tropical forests.^{6,50} They have a smooth tail and different colors, depending on the species and their geographic region. *Bothrops* snakes live in rural areas and in the outskirts of large cities and they prefer humid environments such as forests, plantation areas, and places where there is a proliferation of rodents (eg, bars, silos, and wood deposits). They have nocturnal or crepuscular habits and an aggressive defensive behavior.^{6,12}

Bothrops Venom

Bothrops venom has proteolytic, coagulant, and hemorrhagic activity. A direct nephrotoxic action of the venom also has been shown.

Different venom activities usually are related to the presence of specific components. However, different toxins may have a synergistic activity to induce a particular effect and certain

toxins may have several activities. This variability of toxins and activities also may be observed in different species of *Bothrops*.⁵¹

Local manifestations at the bite site such as edema, blisters, and necrosis are caused by venom proteolytic action. Lesions result from the activity of proteases, esterases, hyaluronidases, and phospholipases (phospholipase A2) released by inflammatory mediators, action of hemorrhagins on the vascular endothelium, and the procoagulant action of the venom.^{6,51,52}

Bothrops venoms activate, either alone or simultaneously, factor X and prothrombin. They also have thrombin-like activity, converting fibrinogen into fibrin. These actions produce coagulation disorders and may lead to blood incoagulability. *Bothrops* venom may induce platelet function abnormalities as well as low platelet count. Hemorrhagic manifestations result from the action of hemorrhagins, metalloproteinases containing zinc, which produce injuries of the capillary basal membranes, associated with low platelet count and coagulation abnormalities. Moreover, hemorrhagins are potent inhibitors of platelet aggregation.^{6,51}

It should be emphasized that for the same species, the composition of venom may vary according to the animal's age (young *B jararaca* and *B moojeni* venoms have greater procoagulant activity and lower local inflammatory activity compared with venom from adult snakes), geographic distribution, and individual characteristics.^{26,29,53-56}

Clinical Manifestations

The clinical picture usually is characterized by early and progressive pain and edema at the bite site. Bruises, blisters, and bleeding also frequently are observed at the venom inoculation area. In most severe cases, there is necrosis of soft tissues with abscess formation and the development of compartmental syndrome, which may result in functional or anatomic loss of the bitten limb.^{6,29} Systemic manifestations include bleeding (pre-existing skin injuries, gingival bleeding, epistaxis, hematemesis, and hematuria), nausea, vomiting, sudoresis, and hypotension. The most severe systemic complications are shock, AKI, septicemia, and disseminated intravascular coagulation-like syndrome.^{6,30,57}

Prevalence of AKI

The reported prevalence of AKI after *Bothrops* envenomation ranges from 1.6% to 38.5% (Table 2). All studies were retrospective and none used creatinine clearance (calculated or measured) or a more sensitive method for the study of renal function.^{32,58-65}

Pathophysiology of AKI

Etiopathogeny of AKI associated with *Bothrops* envenomation has been related to hemodynamic changes, myoglobinuria, hemoglobinuria, coagulation abnormalities, and venom direct nephrotoxicity.^{8,31,66}

Table 2. AKI Prevalence After *Bothrops* Envenomation

Study	Type	n	AKI Diagnosis	% AKI
Cupo et al, 1985 ⁵⁸	Retrospective	67	Creatinine increase	10.5
Pinto et al, 1987 ³²	Retrospective	616	Creatinine increase	1.6
Queiroz and Moritz, 1989 ⁵⁹	Retrospective	114	Method unknown	6
Kouyoumdjian et al, 1990 ⁶⁰	Retrospective	57	Creatinine increase	6
Nishioka and Silveira, 1992 ⁶¹	Retrospective	292	Creatinine increase	5
Ribeiro and Jorge, 1997 ⁶²	Retrospective	3,139	Creatinine increase	1.6
Rodríguez Acosta et al, 2000 ⁶³	Retrospective	60	Method unknown	1.6
Bucaretychi et al, 2001 ⁶⁴	Retrospective	73	Creatinine increase (children <15 y)	1.4
Otero et al, 2002 ⁶⁵	Retrospective	39	Method unknown	38.5

The development of hypotension or shock is a rare event after a *Bothrops* snakebite. Venom may cause hemodynamic abnormalities as a result of sequestration of fluids at the bite site, bleeding, and release of vasoactive substances. It should be noted that the administration of antivenom may cause hypotension or shock as a result of a hypersensitivity reaction.^{31,51,67}

Venom may cause localized muscular injury, but it does not have a systemic myotoxic effect similar to the *Crotalus* venom and it does not induce significant CK increase. Thus, myoglobinuria is unlikely to be an important factor in the pathogenesis of renal injury.^{29,51,68,69}

Bothrops venom is considered hemolytic in vitro and there are clinical reports of anemia and hemolysis after *Bothrops* envenomation, as well as reports of hemoglobinuria after administration of *Bothrops* venom to rats. Hemoglobinuria might contribute to renal injury, worsening renal vasoconstriction, glomerular coagulation, and tubular nephrotoxicity.⁶⁸⁻⁷⁰

Intravenous injection of *B jararaca* venom into rats caused a marked and early decrease of glomerular filtration, renal plasma flow, and diuresis, accompanied by increases in renal vascular resistance and fractional excretion of sodium. There was no hypotension or CK increase. Venom also caused marked fibrinogen consumption and intravascular hemolysis. Renal histologic analysis showed an extensive intraglomerular deposition of fibrin thrombi and acute tubular necrosis.³¹

In fact, Boer-Lima et al^{67,71} studied renal abnormalities induced by *B moojeni* venom in rats and observed a significant decrease of glomerular filtration, occurring in absence of hypotension, and the development of acute tubular necrosis. Later, the same group showed that *B moojeni* venom caused glomerular injury including mesangiolytic, microaneurysms, and glomerular basal membrane abnormalities associated with proteinuria.

In an isolated perfused rat kidney model, it was shown that *B jararaca* venom caused direct acute tubular nephrotoxicity and platelet activating factor might be involved.⁷² Indeed, Castro et al⁶⁶ showed that the *B jararaca* venom caused in vitro injury in isolated renal proximal tubules and *Bothrops* antivenom was

effective only in preventing injury when administered simultaneously with venom.

Risk Factors for the Development of AKI After *Bothrops* Snakebite

Several risk factors have been related to *Bothrops* venom-induced AKI, such as patient age, snake size, species, amount of injected venom, time interval between the bite and administration of the antivenom, and the amount and route of the antivenom administered.⁶⁹ A positive correlation between the patient's age and AKI prevalence has been reported.⁶¹ A retrospective clinical-epidemiologic study performed in Colombia with 39 victims of poisoning by *B asper* showed that the time interval between the accident and administration of the antivenom of more than 2 hours was associated with the development of AKI, as well as with the risk of death and permanent injuries.⁶⁵

Characteristics of AKI

Renal dysfunction after *Bothrops* poisoning occurs early, usually is severe and oliguric, with the need for dialysis varying from 33% to 75% of cases.^{26,47,69} The most frequent renal structural injury found is acute tubular necrosis, although cases of bilateral cortical necrosis, interstitial nephritis, and acute glomerulonephritis with mesangial proliferation also have been reported. Mortality rate of *Bothrops* venom-induced AKI range from 13% to 19%.^{26,31,46,47,65,67,70,71}

REFERENCES

1. Chippaux JP. Snakebites: appraisal of the global situation. Bull World Health Organ. 1998;76:515-24.
2. Barraviera B. Estudo clínico dos acidentes ofídicos. Revisão. J Bras Med. 1993;65:209-50.
3. Pugh RN, Theakston RDG. Incidence and mortality on snakebite in savanna Nigeria. Lancet. 1980;2:81-3.
4. Sutherland SK, Leonard RL. Snakebite deaths in Australia 1992-1994 and management update. Med J Aust. 1995;163:616-8.
5. Nelson BK. Snake envenomation. Incidence, clinical presentation and management. Med Toxicol Adver Drug Exp. 1989;4:17-31.
6. Amaral CFS, Bucarechi F, Araújo FAA, Cardoso JLC, Campos JA, Azevedo-Marques MM, et al. Brasil. Manual de diagnóstico e tratamento de acidentes por

- animais peçonhentos. Brasília: Ministério da Saúde, Fundação Nacional de Saúde; 2001.
7. Sitprija V, Boonpucknavig V. The kidney in tropical snakebite. *Clin Nephrol.* 1977;8:377-83.
 8. Chugh KS. Snake-bite-induced acute renal failure. *Kidney Int.* 1989;35:891-907.
 9. Hoge AR, Romano HS. Sinopse das serpentes peçonhentas. *Mem Inst Butantan.* 1978-1979;42/43:373-496.
 10. Campbell JA. Rattlesnakes, *Crotalus*. In: Campbell JA, Lamar WW, editors. *The venomous reptiles of Latin America*. New York: Cornell University Press Ithaca; 1989. p. 333-45.
 11. Azevedo-Marques MM, Ferreira DB, Costa RS. Rhabdomyonecrosis experimentally induced in wistar rats by Africanized bee venom. *Toxicon.* 1992;30:344-8.
 12. Melgarejo AR. Serpentes peçonhentas do Brasil. In: Cardoso JLC, França FOS, Wen FH, Malaque CMS, Haddad VJ, editors. *Animais peçonhentos no Brasil—biologia, clínica e terapêutica dos acidentes*. São Paulo: Sarvier; 2003. p. 33-62.
 13. Marques OAV, Sazima I. História natural da serpentes. In: Cardoso JLC, França FOS, Wen FH, Malaque CMS, Haddad VJ, editors. *Animais peçonhentos no Brasil—biologia, clínica e terapêutica dos acidentes*. São Paulo: Sarvier; 2003. p. 62-71.
 14. Bercovici D, Chudzinski AM, Esteves MI, Picarelli ZP, Raw I, Ueda CPM, et al. A systematic fractionation of *Crotalus durissus terrificus* venom. *Mem Inst Butantan.* 1987;49:69-78.
 15. Vital BO. Venenos ofídicos neurotóxicos. *Rev Ass Med Bras.* 1980;26:212-8.
 16. Oshima-Franco Y, Hyslop S, Prado JF, Cruz MAH, Rodríguez LS. Neutralizing capacity of antisera raised in horses and rabbits against *Crotalus durissus terrificus* (South American rattlesnake) venom and its main toxin, crotoxin. *Toxicon.* 1999;37:1341-57.
 17. Vital BO, Franceschi JP, Waishich E. Pharmacology of crystalline crotoxin. *Toxicity.* *Mem Inst Butantan.* 1966;33:973.
 18. Azevedo-Marques MM, Cupo P, Coimbra TM, Hering SE, Rossi MA, Laure CJ. Myonecrosis, myoglobinuria and acute renal failure induced by South American rattlesnake (*Crotalus durissus terrificus*) envenomation in Brazil. *Toxicon.* 1985;23:631-6.
 19. Azevedo-Marques MM, Hering SE, Cupo P. Evidence that *Crotalus durissus terrificus* (South American rattlesnake) envenomation in humans causes myolysis rather than hemolysis. *Toxicon.* 1987;25:1163-8.
 20. Cupo P, Azevedo-Marques MM, Hering SE. Clinical and laboratory features of South American rattlesnake (*Crotalus durissus terrificus*) envenomation in children. *Trans R Soc Trop Med Hyg.* 1988;82:924-9.
 21. Salvini TF, Amaral AC, Miyabara EH, Turri JAO, Daneila PM, Araújo HSS. Systemic skeletal muscle necrosis induced by crotoxin. *Toxicon.* 2001;39:1141-9.
 22. Monteiro HAS, Silva IMSC, Martins AMC, Fonteles MC. Actions of *Crotalus durissus terrificus* venom and crotoxin on the isolated rat kidney. *Braz J Med Biol Res.* 2001;34:1347-52.
 23. Martins AMC, Toyama MH, Havt A, Novello JC, Marangoni S, Fonteles MC, et al. Determination of *Crotalus durissus cascavella* venom components that induce renal toxicity in isolated rat kidney. *Toxicon.* 2002;40:1165-71.
 24. Amora DN, Sousa TM, Martins AM, Barbosa PS, Magalhães MR, Toyama MH, et al. Effects of *Crotalus durissus collilineatus* venom in the isolated rat kidney. *Toxicon.* 2006;47:260-4.
 25. Barrio A. Gyroxin, a new neurotoxin of *Crotalus durissus terrificus* venoms. *Acta Physiol Lat Am.* 1961;11:224.
 26. Amaral CFS, Rezende NA, Silva OA, Ribeiro MMF, Magalhães RA, Reis RJ, et al. Insuficiência renal aguda secundária a acidentes ofídicos botrópico e crotálico. Análise de 63 casos. *Rev Inst Med Trop São Paulo.* 1986;28:220-7.
 27. Magalhães RA, Ribeiro MMF, Rezende NA, Amaral CFS. Rabdomiólise secundária a acidente ofídico crotálico (*Crotalus durissus terrificus*). *Rev Inst Med Trop São Paulo.* 1986;28:28-33.
 28. Jorge MT, Ribeiro LA. Epidemiologia e quadro clínico do acidente por cascavel sul americana (*Crotalus durissus*). *Rev Inst Med Trop São Paulo.* 1992;34:347-54.
 29. Jorge MT, Ribeiro LA. Acidentes por serpentes peçonhentas do Brasil. *Rev Assoc Med Bras.* 1990;36:66-77.
 30. Ribeiro LA, Albuquerque MJ, Campos VAFP, Katz G, Takaoka NY, Lebrão ML, et al. Óbitos por serpentes peçonhentas no Estado de São Paulo: avaliação de 43 casos, 1988/93. *Rev Ass Med Bras.* 1998;44:312-8.
 31. Burdmann EA, Woronik V, Prado EB, Abdulkader RC, Saldanha LB, Barreto OC, et al. Snakebite-induced acute renal failure: an experimental model. *Am J Trop Med Hyg.* 1993;48:82-8.
 32. Pinto RNL, Souza LCS, Silva AM, Pereira LIA, Andrade JG. Estudo clínico-epidemiológico de 774 casos de acidentes ofídicos. *Rev Soc Bras Med Trop.* 1987;20 Suppl:56.
 33. Silveira PVP, Nishioka AS. South American rattlesnake bite in a Brazilian teaching hospital. Clinical and epidemiological study of 87 cases, with analysis of factors predictive of renal failure. *Trans Royal Soc Trop Med Hyg.* 1992;86:562-4.
 34. Bucarechi F, Herrera SRF, Hyslop S, Baracat ECE, Vieira RJ. Snakebites by *Crotalus durissus ssp* in children in Campinas, São Paulo, Brazil. *Rev Inst Med Trop São Paulo.* 2002;44:133-8.
 35. Pinho FMO, Zanetta DM, Burdmann EA. Acute renal failure after *Crotalus durissus* snakebite: a prospective survey on 100 patients. *Kidney Int.* 2005;67:659-67.
 36. Vidal EC, Yu L, Castro I, Ori M, Malheiros DM, Burdmann EA. Snake venom-induced nephrotoxicity—in vivo and in vitro studies. *J Am Soc Nephrol.* 1997;8:131A.
 37. Martins AMC, Monteiro HSA, Júnior EOG, Menezes DB, Fonteles MC. Effects of *Crotalus durissus cascavella* venom in the isolated rat kidney. *Toxicon.* 1998;36:1441-50.

38. Sullivan JB, Wingert WA. Reptile bites. In: Averbach PS, Geehr EC, editors. Management of wilderness and environmental emergencies. 2nd ed. St Louis: CV Mosby; 1989. p. 479-511.
39. Amaral CFS, Campolina D, Dias MB, Bueno CM, Chavez C, Penaforte CL, et al. Time factor in the detection of circulating whole venom and crotoxin and efficacy of antivenom therapy in patients envenomed by *Crotalus durissus*. *Toxicon*. 1997;35:699-704.
40. Castro I. *Estudo da toxicidade das peçonhas crotálica e botrópica em túbulos proximais isolados de rins de rato* [doctorate thesis]. São Paulo: Escola Paulista de Medicina, Universidade Federal de São Paulo; 2003. p. 92.
41. Better OS, Stein JH. Early management of shock and prophylaxis of acute renal failure in traumatic rhabdomyolysis. *N Engl J Med*. 1990;322:825-8.
42. Vanholder R, Sever MS, Ereik E, Lameire N. Rhabdomyolysis. *J Am Soc Nephrol*. 2000;11:1553-61.
43. Cupo P, Azevedo-Marques MM, Hering SE. Acidente crotálico na infância: aspectos clínicos, laboratoriais, epidemiológicos e abordagem terapêutica. *Rev Soc Bras Med Trop*. 1991;24:87-96.
44. Zager RA. Rhabdomyolysis and myohemoglobinuric ARF. *Kidney Int*. 1996;49:314-26.
45. Weber RA, White RR. *Crotalidae* envenomation in children. *Ann Plast Surg*. 1993;31:141-5.
46. Silva AO, Lopez M, Godoy P. Intensive care unit treatment of acute renal failure following snakebite. *Am J Trop Med Hyg*. 1979;28:401-7.
47. Vêncio D. Estudo do ofidismo em Goiás: comprometimento da função renal. *Rev Goiana Med*. 1988;34:95-116.
48. Amorim MF, Mello RF, Saliba F. Lesões renais induzidas experimentalmente no cão pelo veneno crotálico. *Mem Inst Butantan*. 1969;34:137-57.
49. Burdmann EA, Barcellos MA, Cardoso JL, Malheiro P, Abdulkader R, Daher E, et al. Acute interstitial nephritis after snake bite. *Ren Fail*. 1989;11:51-2.
50. Greene HW. Snakes. The evolution of mystery in nature. Berkeley: University of California Press; 1997. p. 351.
51. França FOS, Malaque CMS. Acidente botrópico. In: Cardoso JLC, França FOS, Wen FH, Malaque CMS, Haddad VJ. Animais peçonhentos no Brasil—biologia, clínica e terapêutica dos acidentes. São Paulo: Sarvier; 2003. p. 72-86.
52. Machado Braga MD, Costa Martins AM, Alves CD, Menezes DB, Martins RD, Ferreira Barbosa PS, et al. Purification and renal effects of phospholipase A(2) isolated from *Bothrops insularis* venom. *Toxicon*. 2008;51:181-90.
53. Furtado MF. Contribuição ao estudo do veneno de *Bothrops moojeni* em função da idade das serpentes. [doctorate thesis]. São Paulo: Universidade de São Paulo; 1987.
54. Daltry JC, Wuster W, Thorpe RS. Diet and snake venom evolution. *Nature*. 1996;379:537-40.
55. Kamiguti AS, Cardoso JL. Haemostatic changes caused by the venoms of South American snakes. *Toxicon*. 1989;27:955-63.
56. Kamiguti AS, Slupsky JR, Zuzel M, Hay CR. Properties of fibrinogen cleaved by Jararhagin, a metalloproteinase from the venom of *B. jararaca*. *Thromb Haemost*. 1994;72:244-9.
57. Kamiguti AS, Cardoso JL, Theakston RD, Sano-Martins IS, Hutton RA, Rugmann FP, et al. Coagulopathy and hemorrhage in human victims of *Bothrops jararaca* envenoming in Brazil. *Toxicon*. 1991;29:961-72.
58. Cupo P, Azevedo-Marques MM, Hering SE, Menezes JB. Acidentes ofídicos: análise de 102 casos. In: XXI Congresso da Sociedade Brasileira de Medicina Tropical, São Paulo, 1985 Feb 8. Livro de resumos. São Paulo, 1985. p. 23-4.
59. Queiroz LP, Moritz RD. Acidente botrópico em Florianópolis. *Arq Catarinenses Med*. 1989;18:163-6.
60. Kouyoumdjian JA, Polizelli C, Lobo SMA. Acidentes ofídicos causados por *Bothrops moojeni* na Região de São José do Rio Preto-SP. *Arq Bras Med*. 1990;64:167-71.
61. Nishioka AS, Silveira PV. A clinical and epidemiological study of 292 cases of lance-headed viper bite in a Brazilian teaching hospital. *Am J Trop Med Hyg*. 1992;47:805-10.
62. Ribeiro LA, Jorge MT. Acidente por serpente do gênero *Bothrops*: série de 3.139 casos. *Rev Soc Bras Med Trop*. 1997;30:475-80.
63. Rodriguez AA, Uzategui W, Azuaje R, Aguilar I, Giron ME. A clinical and epidemiological analysis of accidental bites by snakes of the genus *Bothrops* in Venezuela. *Rev Cubana Med Trop*. 2000;52:90-4.
64. Bucarechi F, Herrera SRF, Hyslop S, Bcarat ECE, Vieira RJ. Snakebites by *Bothrops* spp in children in Campinas, São Paulo, Brazil. *Rev Inst Med Trop S Paulo*. 2001;43:329-33.
65. Otero R, Gutiérrez J, Beatriz Mesa M, Duque E, Rodríguez O, Luis Arango J, et al. Complications of *Bothrops*, *Portibidium*, and *Bothriechis* snakebites in Colombia. A clinical and epidemiological study of 39 cases attended in a university hospital. *Toxicon*. 2002;40:1107-14.
66. Castro I, Burdmann EA, Seguro AC, Yu L. *Bothrops* venom induces direct renal tubular injury: role for lipid peroxidation and prevention by antivenom. *Toxicon*. 2004;43:833-9.
67. Boer-Lima PA, Gontijo JA, Cruz-Hofling MA. Histologic and functional renal alterations caused by *Bothrops moojeni* snake venom in rats. *Am J Trop Med Hyg*. 1999;61:698-706.
68. Amaral CFS, Da Silva AO, Godoy P, Miranda D. Renal cortical necrosis following *Bothrops jararaca* and *B. jararacussu* snake bite. *Toxicon*. 1985;23:877-85.
69. Burdmann EA, Cais A, Vidal EC. Insuficiência renal aguda nefrotóxica: animais peçonhentos. In: Schor N, Boim MA, Santos OFP, editors. Insuficiência renal aguda: fisiopatologia, clínica e tratamento. São Paulo: Sarvier; 1997. p. 135-41.

70. Rezende NA, Amaral CF, Bambirra EA, Lachatt JJ, Coimbra TM. Functional and histopathological renal changes induced in rats by *B. jararaca* venom. *Braz J Med Biol Res.* 1989;22:407-16.
71. Boer-Lima PA, Gontijo JA, Cruz-Hofling MA. *Bothrops moojeni* snake venom-induced renal glomerular changes in rat. *Am J Trop Med Hyg.* 2002;67:217-22.
72. Monteiro HAS, Fonteles MC. The effect of *B. jararaca* venom on rat kidney after short-term exposure: preliminary results. *Pharmacol Toxicol.* 1999;85:198-200.