

# Serum allergen specific immunoglobulin E levels in patients with allergic conjunctivitis

S. KOCABEYOGLU<sup>1</sup>, B. BOZKURT<sup>2</sup>, O. BILEN<sup>3</sup>, M. IRKEC<sup>1</sup>, M. ORHAN<sup>1</sup>

<sup>1</sup>Department of Ophthalmology, Hacettepe University School of Medicine, Ankara

<sup>2</sup>Department of Ophthalmology, Selcuk University, Meram Medical Faculty, Konya

<sup>3</sup>Kudret Eye Hospital, Ankara - Turkey

**PURPOSE.** *To evaluate serum allergen specific immunoglobulin E (IgE) levels in patients with various types of allergic conjunctivitis.*

**METHODS.** *Twenty-five patients with seasonal allergic conjunctivitis (SAC), 17 patients with perennial allergic conjunctivitis (PAC), and 10 patients with vernal conjunctivitis (VC) were included in the study. Specific IgE levels to Dermatophagoides pteronyssinus (Dp), Dermatophagoides farinae (Df), mixed grass pollens, and animal epithelia were measured using Pharmacia CAP system (Pharmacia Diagnostic AB, Uppsala, Sweden).*

**RESULTS.** *The percentage of subjects with specific IgE against Dp and Df was statistically higher in VC (30%) compared to PAC (5.9%) and SAC (0%) ( $p=0.03$ ). Specific IgE against mixed grass pollens was found in 30% of VC and 40% of SAC, whereas 10% of VC and 8% of SAC patients were found to be hypersensitive to animal epithelia.*

**CONCLUSIONS.** *Allergic reaction against house dust mites and pollens was common in VC, whereas specific IgE against grass pollens was remarkable in SAC. IgE levels specific to various antigens might be measured by UNICAP system, which is a rapid and practical technology. (Eur J Ophthalmol 2008; 18: 675-9)*

**KEY WORDS.** *Allergic conjunctivitis, Serum specific IgE, UNI-CAP system*

*Accepted: February 20, 2008*

## INTRODUCTION

Allergic reaction can occur at various sites, including the skin, respiratory tract, and eye, with accompanying symptoms ranging from mildly irritating to severe and chronic. Conjunctiva is one of the most frequently affected sites in an allergic reaction. There are various types of allergic conjunctivitis, including seasonal allergic conjunctivitis (SAC), perennial allergic conjunctivitis (PAC), vernal conjunctivitis (VC), atopic conjunctivitis (AC), and giant papillary conjunctivitis (GPC) (1-3). The incidence of ocular allergy varies markedly among different geographic regions, being more likely in individuals living in warm climates. SAC and PAC represent the majority of ocular allergic disorders. They are caused by direct exposure of the ocular

mucosal surfaces to the allergens, followed by reaction of these allergens with specific immunoglobulin (Ig) E antibodies bound to the surface of conjunctival mast cells resulting in degranulation and release of chemical mediators, including histamine, prostaglandins, leukotrienes, tryptase, and cytokines (4-7). SAC is most commonly related to atmospheric pollens, whereas PAC is related to animal dander, dust mites, or other environmental year-round allergens (8, 9). VC is a rare condition which typically develops in childhood and is more common in males (10, 11). Previous studies (12, 13) have demonstrated increased IgE levels specific for house dust, dust mites, fungi, animal epithelia, and grass antigen in patients with VC. As allergic conjunctivitis is a growing social concern worldwide, it is crucial to detect the causal allergens to

prevent and treat the disease. The purpose of this study is to determine the serum allergen specific IgE levels in subjects with various allergic conjunctivitis using Pharmacia CAP system (Pharmacia Diagnostic AB, Uppsala, Sweden).

## MATERIALS AND METHODS

Patients with signs and symptoms of allergic conjunctivitis were recruited into the study from March to September. The procedures used in this study conformed to the tenets of the Declaration of Helsinki and the Ethics Committee of Hacettepe University Faculty of Medicine approved the research. Informed consent was obtained from the participating individuals following an explanation of the nature and possible consequences of the study. A complete eye examination including visual acuity testing, refraction, slit-lamp biomicroscopic examination, and fundus examination was performed. Demographic data, including age at inclusion, age at onset, gender, association with concomitant or previous systemic allergic conditions, medications, and family history were evaluated.

The diagnostic criteria of SAC patients were bilateral involvement, acute attack, watery discharge, itching, papillary hypertrophy, and alteration of symptoms with seasons (14), whereas PAC occurs throughout the year. The main features of VC are itching, photophobia, mucoid discharge, and giant papillae on the upper tarsal conjunctiva. VC was classified as palpebral if the patient had giant papillae on the upper tarsal conjunctiva, as limbal if the patient had limbal papillae and Trantas dots, and as mixed if the patient had both palpebral and limbal signs. Patients with blepharitis and contact lens wearers were excluded from the study.

Twenty-five patients (17 male, 8 female) with SAC, 17 patients (7 male, 10 female) with PAC, and 10 patients (8 male, 2 female) with VC, who had been treated at the Hacettepe University School of Medicine, Department of Ophthalmology, were included in the study. The mean ages of subjects were  $13.5 \pm 4.6$  years,  $19.2 \pm 13.53$  years, and  $18.2 \pm 12.6$  years in VC, SAC, and PAC groups, respectively ( $p=0.56$ ). In the VC group, 2 patients had limbal VC, 6 patients had palpebral VC, and 2 patients had mixed type VC.

Ten milliliters peripheral blood was taken from the antecubital vein and centrifuged, and serum samples were collected and stored at  $-20^{\circ}\text{C}$ . UniCAP specific IgE assay is

an in vitro quantitative assay used for the measurement of allergen specific IgE in human serum or plasma. UniCAP specific IgE assay is used with the instrument ImmunoCAP 100/UniCAP 100. The allergen of interest, covalently coupled to ImmunoCAP, reacts with the specific IgE in the patient sample. After washing away non-specific IgE, enzyme-labeled antibodies against IgE (enzyme-anti-IgE) are added to form a complex. After incubation, unbound enzyme-anti-IgE is washed away and the bound complex is then incubated with a developing agent. After stopping the reaction, the fluorescence is measured. The specific IgE assay was calibrated according to WHO standard (standard WHO 75/502), in the range of 0.35–100 kilounits per liter (kU/L) and the levels were read from a standard curve. According to Pharmacia CAP system instructions, a value greater than 0.35 kU/L was defined as a positive result. The values 0.35–0.69 kU/L were grouped in class 1, 0.70–3.49 kU/L in class 2, 3.50–17.49 kU/L in class 3, 17.5–49.99 kU/L in class 4, and 50.0–100 kU/L in class 5. One kU/L corresponds to 2.4 ng IgE/mL. In all patients specific IgE levels against grass pollen (mixed grass pollen, gx1, g3, g4, g5, g6, g8; *Dactylis glomerata*, *Festuca elatior*, *Lolium perenne*, *Phleum pratense*, *Poa pratensis*), house dust mites (*Dermatophagoides pteronyssinus* [dp, d1], *Dermatophagoides farinae* [df, d2]), and mixed animal epithelia (ex1-cat, horse, cow, dog dander) were measured.

In statistical analysis, age at inclusion was compared using Student *t*-test, while gender, family history, associated allergic diseases, and positive IgE against allergens were analyzed using chi-square test and Fisher exact test and a *p* value  $< 0.05$  was considered as statistically significant.

## RESULTS

The age at onset was before 10 years in 70% of VC, 72% of SAC, and 64.7% of PAC subjects ( $p>0.05$ ). Family history was present in 80% of VC, 58.8% of PAC, and 32% of SAC ( $p=0.03$ ). The percentages of associated allergic rhinitis, asthma, and atopic dermatitis in each group can be seen in Table I. There were no statistically significant differences in the incidence of these diseases among the three groups ( $p>0.05$ ).

The percentage of subjects with specific IgE against dp and df was statistically higher in VC (30%) compared to PAC (5.9%) and SAC (0%) ( $p=0.03$ ). Specific IgE against

mixed grass pollens was found in 30% of VC, 40% of SAC, and none of the subjects with PAC, whereas specific IgE against animal epithelia was found in 10% of VC, 8% of SAC, and none of the subjects with PAC. The distribution of positive subjects according to five classes is given in detail in Table II. In subjects with associated allergic disorders, the IgE positivity was 33.3% in VC group, 40% in SAC group, and 20% in PAC group.

## DISCUSSION

The diagnosis of allergic conjunctivitis is mainly based on history and clinical examination. Laboratory tests may be useful to confirm the clinical diagnosis (15) and determination of serum IgE levels is one of the useful tests. UNICAP system is a solid-phase immunoassay, used for the titration of serum specific IgE. Ewan et al (16) compared the Pharmacia CAP system and radioallergosorbent test (Phadebas RAST) for five inhalant allergens, dp, timothy grass pollen, cat epithelium, *Cladosporium*, and *Alternaria*. They found that the CAP system was more sensitive than the RAST. In another study by Bousquet et al (17), the sensitivity and the specificity of CAP were found better than RAST except for orchard-grass pollen. Boccagni et al (18) compared four techniques (CAP Sys-

tem, Kallested Allercoat System, Neo Abello, Hamlet-IgE) for specific IgE detection and they showed that the CAP system offers the highest sensitivity. Pastorello et al (19) found high sensitivity (95.5%) and specificity (98.1%) for the CAP system.

SAC symptoms become more frequent and severe with increasing pollen counts (20-23). Mimura et al (24) evaluated serum allergen specific IgE levels against frequent allergens in Japan (five animal epithelia, house dust, *Dermatophagoides pteronyssinus*, *Acarus*, moth, *Candida*, *Alternaria tenius*, Japanese cypress pollen, ragweed, mugwort, and orchard grass) using UNICAP. In SAC group, specific IgE positivity against cedar pollen, cypress pollen, dp, and animal epithelia were 52.5%, 37.5%, 30%, and 5%, respectively (24). In another study, Mimura et al (25) found IgE positivity against house dust and dp in 51.9% and 48.1%, respectively, in autumn group and cedar (68.8%) and cypress pollen (59.4%) in the spring group. There were studies from Turkey about pollen types from different regions (26-29). Arboreal pollen types are predominant in the atmosphere of Bursa, Balikesir, and Ankara. In their study, Pinar et al (27) found high pollen concentration in June and *Pinaceae*, *Cupressaceae/Taxaceae*, *Gramineae*, *Platanus*, *Populus*, *Moraceae*, *Chenopodiaceae/Amaranthaceae*, *Acer*, *Quercus*, *Betula*, *Salix*, *Rumex*, and *Plantago* were found to be the dominant pollen types in Ankara, Turkey.

Grass pollens are known as the major allergens worldwide, especially in the temperate zone (20, 30). The most important pollens are grasses and olive trees in Spain from April to June (31), grass pollens in Greece from March to June (32), and grass pollens in summer in Poland (30). In this study, we evaluated specific IgE positivity against mixed grass pollen (gx1) in allergic subjects in a period between May and September. In our SAC group, positive IgE against grass pollens was observed in

**TABLE I - THE PERCENTAGE OF ASSOCIATED ALLERGIC DISORDERS**

	Seasonal allergic conjunctivitis	Perennial allergic conjunctivitis	Vernal conjunctivitis
Rhinitis	28.2%	17.6%	20%
Asthma	12.2%	11.7%	20%
Dermatitis	4%	11.7%	10%

**TABLE II - SERUM-SPECIFIC IGE LEVELS AGAINST VARIOUS ANTIGENS**

	SAC			PAC			VC		
	dp, df	Grass	Animal epithelia	dp, df	Grass	Animal epithelia	dp, df	Grass	Animal epithelia
Class I	0	1	1	0	0	0	0	0	0
Class II	0	1	1	1	0	0	3	0	1
Class III	0	5	0	0	0	0	0	2	0
Class IV	0	3	0	0	0	0	0	1	0
Class V	0	0	0	0	0	0	0	0	0

40% of the subjects, against animal epithelia in 8% of the subjects, whereas none of the SAC subjects had measurable specific IgE against dp and df. The percentage of SAC patients with positive serum specific IgE was low compared to the literature. We measured only IgE levels against grass pollens; however, some of these patients might be allergic to arboreal pollens. Another limitation of this study was that the diagnosis of the allergic subjects was made according to their clinical symptoms and signs. We did not perform prick tests or conjunctival cytology to all of them. Only patients with associated allergic diseases (asthma, atopic dermatitis) were evaluated by the allergologists and prick tests were performed to these subjects.

Fujishima et al (12) found house dust positivity in 60%, house dust mite positivity in 80%, and cat epithelia in 60% of the 10 subjects with VC. Tuft et al (13) evaluated 120 patients with VC and found that serum specific IgE levels against grass pollen were 90% in the limbal group and 94% in the palpebral and mixed group, which were higher than the control group (29%). The positivity against cat and mite ranged from 82–100% in VC and 14–29% in the control group. Kitazawa et al (33) found IgE positivity against Japanese cedar pollen as 34.3% in SAC group, against house dust mites as 81.8% in VC, and 40% in PAC groups. In our VC group, specific IgE positivity against house dust mite, mixed grass pollen, and animal epithelia were 30%, 30%, and 10%, respectively, which were lower than the previous studies. The number of subjects was very low and we have to enlarge our sample size to confirm these results in VC group.

Gradman et al (34) found allergic conjunctivitis in 42% of the subjects with allergic rhinitis, 24% of the subjects with asthma, and 30% of the subjects with dermatitis. Sensitization to house dust mites was more frequent in chronic allergic conjunctivitis than in acute allergic conjunctivitis

(95% vs 53%) and sensitization to grass pollen was more frequent in children with acute allergic conjunctivitis (34). Leonardi et al (35) found associated allergic diseases in 46% of the VC cases and the most frequent disease was rhinitis (30.1%), followed by eczema (16.3%) and asthma (14.6%). In a case series of 109 patients with VC from Nigeria (36), a history of atopic diseases was present in only 4.5% of the subjects. In our study, allergic rhinitis was the most common concomitant allergic disease in VC (20%) and SAC (28%). As ocular allergy might be a part of a systemic allergic disorder such as rhinitis, asthma, and/or dermatitis, these subjects should be managed in collaboration with an allergologist.

Ocular allergic diseases might occur as a result of sensitization to various antigens, such as pollens, house dust mites, and animal epithelia, which might be detected simply by measuring specific IgE levels using UNICAP system, a rapid and easy technology. The allergens should better be selected according to the region of the country and the type of the ocular allergic disorder. Specific IgE measurement to only limited antigens is not always a sensitive diagnostic test in ocular allergic diseases, since it might be negative in almost half of the subjects. However, as shown in our study, these tests might be helpful to determine the extent of sensitization of the subjects to selected antigens and might aid in the comprehensive management of the problem.

*Proprietary interest: None.*

Reprint requests to:  
Murat Irkeç, MD  
Hacettepe Üniversitesi  
Göz Hastalıkları Anabilim Dalı  
Sıhhiye, 06100  
Ankara, Turkey  
mirkeç@isnet.net.tr

---

## REFERENCES

1. Abelson MB, Schaefer K. Conjunctivitis of allergic origin: immunologic mechanisms and current approaches to therapy. *Surv Ophthalmol* 1993; 38: 115.
2. Friedlander MH. Conjunctivitis of allergic origin: clinical presentation and differential diagnosis. *Surv Ophthalmol* 1993; 38: 105.
3. Allansmith MR, Korb DR, Greiner JV, Henriquez AS, Simon MA, Finnemore VM. Giant papillary conjunctivitis in contact lens wearers. *Am J Ophthalmol* 1977; 83: 697-708.
4. McGill JI, Holgate ST, Church MK, Anderson DF, Bacon A. Allergic eye disease mechanisms. *Br J Ophthalmol* 1998; 82: 1203-14.
5. Anderson DF, Zhang S, Bradding P, McGill JI, Holgate ST, Roche WR. The relative contribution of Mast cell subsets to conjunctival Th2 like cytokines. *Invest Ophthalmol and Vis Sci* 2001; 42: 995-1001.

6. Bielory L. Allergic and immunologic disorders of the eye. Part II: Ocular allergy. *J Allergy Clin Immunol* 2000; 106: 1019-32.
7. Abelson MB, Allansmith MR. Histamine in the eye. In: Silverstein A, O'Connor G, eds. *Immunology and Immunopathology of the Eye*. New York: Masson Publishing, 1979; 362-4.
8. Mimura T, Yamagami S, Amano S, et al. allergens in Japanese patients with allergic conjunctivitis in autumn. *Eye* 2005; 19: 995-9.
9. Stahl JL, Barney NP. Ocular allergic disease. *Curr Opin Allergy Clin Immunol* 2004; 4: 455-9.
10. Buckley RJ. Vernal keratoconjunctivitis. *Int Ophthalmol Clin* 1988; 28: 303-8.
11. Allansmith MR. Vernal conjunctivitis. In: Duane TD ed. Vol. 4, Chap. 9. *Clinical Ophthalmology*. New York: Harper and Row, 1980; 6.
12. Fujishima H, Saito I, Takeuchi T, Tsubota K. Immunological characteristics of patients with vernal keratoconjunctivitis. *Jpn J Ophthalmol* 2002; 46: 244-8.
13. Tuft SJ, Dart JKG, Kemeny M. Limbal vernal keratoconjunctivitis: clinical characteristics and immunoglobulin E expression compared with palpebral vernal. *Eye* 1989; 3: 420-7.
14. Ono SJ, Abelson MB. Allergic conjunctivitis: update on pathophysiology for future treatment. *J Allergy Clin Immunol* 2005; 115: 118-22.
15. Asbell PA, Ahmad SM. Diagnostic assays in ocular allergy. *Int Ophthalmol Clin* 2003; 43: 83-93.
16. Ewan PW, Coote D. Evaluation of a capsulated hydrophilic carrier polymer (the immunoCAP) for measurement of specific IgE antibodies. *Allergy* 1990; 45: 22-9.
17. Bousquet J, Chanez P, Chanal I, Michel FB. Comparison between RAST and Pharmacia CAP system: a new automated specific IgE assay. *J Allergy Clin Immunol* 1990; 85: 1039-43.
18. Boccagni P, Favari F, Zanoni G, Pezzini A, Tridente G. Comparison of four in vitro assays for specific IgE detection. *Int J Clin Lab Res* 1994; 24: 102-5.
19. Pastorello EA, Incorvaia C, Pravettoni V. A multicentric study on sensitivity and specificity of a new in vitro test for measurement of IgE antibodies. *Ann Allergy* 1991; 67: 365-70.
20. Burr ML, Emberlin CJ, Treu R, Cheng S, Pearce NE, the ISAAC Phase One Study Group. Pollen counts in relation to the prevalence of allergic rhinoconjunctivitis, asthma and atopic eczema in the international study of asthma and allergies in childhood (ISAAC). *Clin Exp Allergy* 2003; 33: 1675-80.
21. Davies RR, Smith LP. Forecasting the start and severity of the hay fever season. *Clin Allergy* 1973; 3: 263-7.
22. Viander M, Koivikko A. The seasonal symptoms of hyposensitized and untreated hayfever patients in relation to birch pollen counts: correlations with nasal sensitivity, prick test and RAST. *Clin Allergy* 1978; 8: 387-96.
23. Koivikko A, Kupius R, Makinen Y, Pohjala N. Pollen seasons forecast of the most important allergenic plants in Finland. *Allergy* 1986; 41: 233-42.
24. Mimura T, Amano S, Funatsu H, et al. Correlations between allergen-specific IgE serum levels in patients with allergic conjunctivitis in spring. *Ocular Immunol Inflamm* 2004; 12: 45-51.
25. Mimura T, Yamagami S, Amano S, et al. allergens in Japanese patients with allergic conjunctivitis in autumn. *Eye* 2005; 19: 995-9.
26. Bicakci A, Akyalcin H. Analysis of airborne pollen fall in Balikesir, Turkey 1996-1997. *Ann Agric Environ Med* 2000; 7: 5-10.
27. Pinar NM, Sakiyan N, Inceoglu Ö, Kaplan A. A one-year aeropalynological study at Ankara, Turkey. *Aerobiologia* 1999; 15: 307-10.
28. Bicakci A, Inceoglu Ö, Sapan N, Malyer H. Airborne pollen calendar of the central region of Bursa (Turkey). *Aerobiologia* 1996; 12: 43-6.
29. Bicakci A, Tatlidil S, Sapan N, Malyer H, Canitez Y. Airborne pollen grains in Bursa, Turkey, 1999-2000. *Ann Agric Environ Med* 2003; 10: 31-6.
30. Puc M, Puc MI. Allergenic airborne grass pollen in Szczecin, Poland. *Ann Agric Environ Med* 2004; 11: 237-44.
31. Subiza Garrido-Lestache J. Allergenic pollens in Spain. *Allergol Immunopathol* 2004; 32: 121-4.
32. Syrigou E, Zanikou S, Papageorgiou PS. Grasses, olive, Parietaria and cypress in Athens: pollen sampling from 1995 to 1999. *Aerobiologia* 2003; 19: 133-7.
33. Kitazawa M, Shoji J, Inada N, Sawa M, Kato H. Clinical evaluation of measurement method of antigen specific IgE in tears of patients with allergic conjunctival disease. *Nippon Ganka Gakkai Zasshi* 2003; 107: 578-82.
34. Gradman J, Wolthers OD. Allergic conjunctivitis in children with asthma, rhinitis and eczema in a secondary outpatient clinic. *Pediatr Allergy Immunol* 2006; 17: 524-6.
35. Leonardi A, Busca F, Motterle L, et al. Case series of 406 vernal keratoconjunctivitis patients: a demographic and epidemiological study. *Acta Ophthalmol Scand* 2006; 84: 406-10.
36. Ukponmwan CU. Vernal keratoconjunctivitis in Nigerians: 109 consecutive cases. *Trop Doct* 2003; 33: 242-5.

Copyright of European Journal of Ophthalmology is the property of Wichtig Editore and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.